



## PGA Bacterial DNA Extraction kit

CatalogNo.PD115-050

50 Preps

Store: RT

Scan to learn how to use



### Kit CONTENTS:

Buffer I	5 ml
Buffer II	10 ml
Buffer III	8 ml
Buffer X	10 ml
Rnase A	50 µl
Solvent Buffer	5 ml

Required contents:

cool ethanol %100  
cool ethanol %70

**Attention: before use, please add RNase A to Buffer I and keep in 4 °C**

### LABORATORY PROTOCOL:

- 1: Collect 3 -5 ml bacterial culture in a microtube, then centrifuge at 13000 rpm for 3 minutes.
- 2: Resuspend the precipitate in 100µl Buffer I.
- 3: Add 200 µl Buffer II in the microtube and gently invert 3-5 times.
- 4: Add 150 µl Buffer III in the microtube and gently invert 3 times, then keep at 37 °C for 45 minutes. (For gram-negative bacteria no need number 4 step)
- 5: Add 180 µl Buffer X in the microtube, then invert 10 times. **The white precipitate should be seen.**
- 6: Microcentrifuge 13000 rpm for 10 minutes.
- 7: Transfer the supernatant to a new microtube. **(Important: do not transfer precipitate in a new tube. otherwise, repeat step 6)**
- 8: Add cold ethanol %96 - %100(2 × supernatant volume) to the supernatant and invert gently 5 times.
- 9: Microcentrifuge at 13000 rpm for 5 minutes.
- 10: Pour off the ethanol by inverting the tube gently, then keep precipitate.
- 11: Wash the precipitate by adding 700µl cold ethanol %70 and invert 2-3 times.
- 12: Microcentrifuge 13000 rpm for 1 minute.
- 13: Pour off the ethanol entirely and dry the pellet for 2-3 minutes at room temperature.
- 14: According to precipitate, Add 20 - 50µl Solvent Buffer in the tube. The precipitate must be solved completely.